

A flexible, multi-week approach to plant biology - How will plants respond to higher levels of CO₂?

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Abstract

To help introductory biology students understand how plants will respond to higher levels of CO₂, we have created a multi-week module consisting of a series of four related laboratory lessons. These lessons allow students to make connections among morphological, physiological, and growth responses of *Brassica rapa* (Fast Plants(R)) to low (400 ppm) and high (800 ppm) concentrations of atmospheric carbon dioxide (CO₂). Elevated CO₂ was chosen as an environmentally relevant variable of high interest to our students as they consider the environment of their futures. However, this module is easily adaptable to test other plant species and/or variables (e.g., light, nutrients, soil contaminants, genotypes, biotic factors, etc.), depending on instructor/student interests and available facilities/equipment. Within our modular framework, students analyze stomatal densities, photosynthetic and respiratory rates, foliar protein concentrations, and growth/allocation patterns. Progressive, weekly analysis of *B. rapa* responses to high and low CO₂ concentrations allows students to apply their findings and make predictions about plant responses in subsequent lessons. After all responses are measured, this framework facilitates student synthesis as they describe the interconnectedness of the measured processes and reflect on how these responses contribute to “how plants will respond to higher levels of CO₂.”

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Supporting Materials: S1. Plants and CO₂-Lesson 1: Stomatal Morphology and Density: Student Lab Manual Pages, S2. Plants and CO₂-Lesson 2: Photosynthesis and Respiration: Student Lab Manual Pages, S3. Plants and CO₂-Lesson 3: Foliar Protein Concentrations: Student Lab Manual Pages, S4. Plants and CO₂-Lesson 4: Growth and Resource Allocation: Student Lab Manual Pages, S5. Plants and CO₂-Supply List per Lesson, S6: Plants and CO₂-Pre-module Background Reading: Plant Responses to CO₂, S7. Plants and CO₂-Group Member Evaluation Form, S8. Plants and CO₂-Example Quiz, S9. Plants and CO₂-Lab Notebook Sample Grading Rubric, and S10. Plants and CO₂-Collaborative Report Instructions and Grading Rubric (.docx).

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Learning Goal(s)

Students will:

- Understand the scientific method, including asking questions, formulating hypotheses, gathering and analyzing data, predicting outcomes, and displaying results.
- Learn how to keep a scientific lab notebook.
- Learn about the components of a scientific paper and develop skills in literature research and scientific writing.
- Develop collaborative skills.
- Understand basic statistical analysis of biological data.
- Gain basic knowledge about plant anatomy/physiology and better understand the relationships between environmental conditions and plants.

Learning Objective(s)

Students will be able to:

- Apply findings from each week's lesson to make predictions and informed hypotheses about the next week's lesson.
- Keep a detailed laboratory notebook.
- Write and peer-edit the sections of a scientific paper, and collaboratively write an entire lab report in the form of a scientific research paper.
- Search for, find, and read scientific research papers.
- Work together as a team to conduct experiments.
- Connect findings and ideas from each week's lesson to get a broader understanding of how plants will respond to higher levels of CO₂ (e.g., stomatal density, photosynthetic/respiratory rates, foliar protein concentrations, growth, and resource allocation).

Note: Additional, more specific objectives are included with each of the four lessons (Supporting Files S1-S4)

INTRODUCTION

Origin and Rationale

In our previous introductory biology course sequence, students engaged in instructive, research-like lessons in hands-on laboratory courses. However, these lessons were relatively unrelated to one another, lacking obvious connections from lesson to lesson. We found that although students learned the scientific process and a wide array of biological topics, their learning remained at a superficial level. We think that this shortcoming reflected the structure of the course, which did not support students' ability to see how these lessons and their respective findings related to one another. Therefore, we redesigned our introductory biology curriculum into modules, containing a series of intentionally related lessons. This paper describes one module, which explores how plants will respond morphologically, physiologically, and biochemically to higher levels of CO₂.

In this four-week module, student groups investigate plants and the connections among stomatal density (Supporting File S1: Stomatal Density Lesson), photosynthetic/respiratory rates (Supporting File S2: Photosynthesis and Respiration Lesson), foliar protein concentrations (Supporting File S3: Foliar Protein Lesson), and growth/resource allocation (Supporting File S4: Growth and Allocation Lesson). The stomatal imprint methodology in Lesson 1: Stomatal Density (Supporting File S1) is adapted from Grant and Vatnink 1998 (1). We specifically analyze how *Brassica rapa* (Fast Plants®) respond to low (400 ppm) and high (800 ppm) concentrations of atmospheric CO₂. Elevated CO₂ was chosen as our independent variable because it is an environmentally relevant and engaging topic as our students consider the environment of their futures. *Brassica rapa* was chosen because it is a fast-growing plant with a short life cycle (i.e., flower two weeks after planting); it is an ideal model organism for short-term studies. Although we employed large growth chambers to control CO₂ concentrations (Supporting Files S1: Stomatal Density Lesson; Supporting File S5: Supply List), the module is easily adaptable to other plant species and/or independent variables (e.g., light, nutrients, soil contaminants, genotypes, temperatures, biotic factors, etc.). Thus, the module can be adapted to instructor/student interests and available facilities/equipment.

Concentrations of atmospheric CO₂ have dramatically risen in recent history (Figure 1) (2), primarily due to combustion of fossil fuels by humans (3). Pre-industrial levels of atmospheric CO₂ were 280 ppm, but current levels are 401 ppm (Figure 1) (2). Many scientists are concerned about this increase because analysis of ancient gas trapped in ice cores indicates that CO₂ concentrations were confined to between 180 and 280 ppm during the past 800,000 years (4,5). In fact, Earth has not experienced such consistently high concentrations of CO₂ since the middle Miocene, approximately 15-20 million years ago (5). Since humans are predicated to continue to burn fossil fuels, CO₂ concentrations are projected to reach 550 ppm by 2050 (6) and could be as high as 1000 ppm by the end of the century (7).

The direct impact of elevated concentrations of atmospheric CO₂ on plant morphology, physiology, biochemistry, and growth has been an active area for research (8,9) and teaching

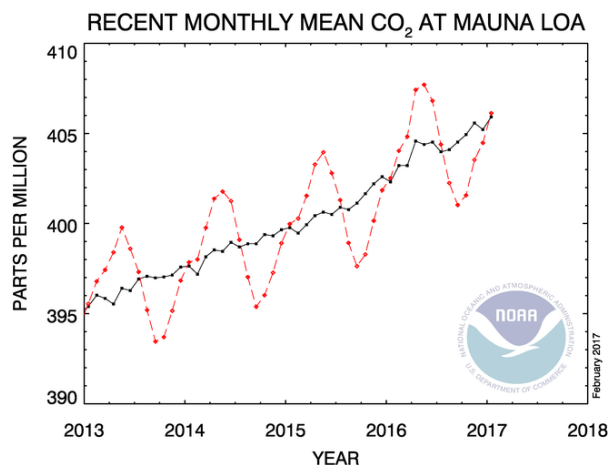


Figure 1. Trends in atmospheric carbon dioxide levels in recent history as recorded by NOAA (1). Used per the US Copyright Act section 105.

(10,11). The focus on CO₂ is not surprising considering that assimilation of CO₂ by plants supports most heterotrophic life on Earth. Plants take in CO₂ (an inorganic compound) from the atmosphere through their stomata via the process of photosynthesis. CO₂ is then used to synthesize organic compounds, which are used by plants for maintenance, growth, and reproduction.

Lesson 1: Stomatal response to elevated CO₂ (Supporting File S1: Stomatal Density Lesson)

All plants do not respond to elevated atmospheric conditions of CO₂ in the same way since like other living organisms, plant responses are influenced by both their genetic make-up and by the environment in which they live. Plants control the rate at which CO₂ enters and H₂O exits stomata by physiologically regulating the size of the stomatal opening or by morphologically modifying stomatal density during leaf expansion (e.g., 12, 13, 14, 15). In our first lesson, we investigate the latter of these plant responses by analyzing the stomatal density of *B. rapa* grown in elevated CO₂ compared to those grown in low CO₂. Reducing water loss from transpiration through stomata is important for many plants to maintain cell turgor and growth. However, minimizing water loss through stomata (via transpiration) can also result in a decrease of CO₂ concentrations inside the plant, which reduces photosynthetic rates and the ability of plants to make carbon-based molecules. Thus, stomata control is an important tradeoff in plants: they must allow sufficient CO₂ into the plant to maintain growth, while minimizing loss of water. This means that for plant species that regulate stomatal conductance by altering stomatal density during leaf expansion, lowering stomatal density in high CO₂ environments will generally increase fitness because plants can take in sufficient amounts of CO₂ with minimal water loss (12,15).

Lesson 2: Photosynthetic and respiratory responses to elevated CO₂ (Supporting File S2: Photosynthesis and Respiration Lesson)

A common physiological response of plants to elevated CO₂ is an increase in photosynthetic rates, which can fuel increased

growth rates and biomass accumulation (8,14). According to a meta-analysis of many scientific studies, plants grown under elevated CO₂ conditions exhibit an average of a 31% increase in photosynthetic rates compared to plants grown under ambient CO₂ conditions (16). This general response of plants, however, is not universal as photosynthetic rates of plants to elevated CO₂ can be depressed by low temperatures and low water availability (17). Additionally, the responsiveness of plant species to elevated CO₂ is strongly regulated by the metabolic pathway used to bind CO₂ during photosynthesis. Photosynthetic rates of C3 plants respond most positively to elevated CO₂ while C4 plants respond to a lesser extent or not at all (14,18).

The processes of photosynthesis and respiration are connected in that sugars made by photosynthesis are used via respiration to provide energy for growth and maintenance. However, there is no clear consensus on how plant respiration responds to elevated CO₂ concentrations as increases, decreases, and no change have been reported (19,20).

Lesson 3: Response of foliar protein concentrations to elevated CO₂ (Supporting File S3: Foliar Protein Lesson)

A common response of plants to elevated CO₂ concentrations is a reduction in nitrogen concentrations in plant tissues. Meta-analyses indicate average reductions in nitrogen concentrations across plant species by approximately 10-15% (8,21). Many mechanisms have been proposed to explain reduced nitrogen concentrations in plants grown in elevated CO₂ conditions. Two likely mechanisms include (1) dilution of nitrogen by increased photosynthetic rates and synthesis of carbon-based molecules and (2) reduction in nitrogen uptake by roots (22). Protein concentrations are linked to nitrogen concentrations in plant tissues. Therefore, it is no surprise that tissues of major food crops express a reduction in protein concentrations of approximately 10-15% (23). Decreases in protein concentrations in leaves are largely due to reductions in Rubisco (rubulose-1,5-bisphosphate carboxylase/oxygenase), the enzyme that binds to CO₂ in the chloroplasts of plants (14).

Lesson 4: Increased plant biomass under elevated CO₂ (Supporting File S4: Growth and Allocation Lesson)

Increased photosynthetic rates and water use efficiency of plants growing in elevated CO₂ conditions typically translates into increased growth rates and biomass accumulation. Meta-analyses suggest an increase of approximately 20-29% in total plant biomass and more than 30% increase in biomass of below ground tissues (8,14).

***Brassica rapa* (Fast Plants®): a model plant**

Due to their rapid rate of maturation and small size, *B. rapa* are commonly utilized as model plants for investigations in elementary, high school, and college classrooms (24). The topics of these investigations are broad, spanning several subdisciplines of plant biology. Classroom experiments on *B. rapa* have been published investigating genetics (25,26,27), plant physiology [nutrition (28,29), tropisms (28), photosynthesis (30)], ecology (31,32), plant development (33), and evolution (34). Previously published activities using *B. rapa* largely focus on a particular topic like Mendelian genetics or artificial selection, whereas our module allows

students to investigate how an environmental factor affects several attributes and processes fundamental to plant growth and survival. Therefore, this module fills a need for a suite of undergraduate-level investigations about *B. rapa* in an environmentally relevant context.

In this module, we investigate how elevated concentrations of CO₂ will affect stomatal density (Supporting File S1: Stomatal Density Lesson), photosynthesis (Supporting File S2: Photosynthesis and Respiration Lesson), respiration (Supporting File S2: Photosynthesis and Respiration Lesson), protein concentrations (Supporting File S3: Foliar Protein Lesson), growth (Supporting File S4: Growth and Allocation Lesson) and resource allocation (Supporting File S4: Growth and Allocation Lesson) in *B. rapa*. For our investigations, 400ppm (low) CO₂ concentration was chosen to model current CO₂ conditions. We used 800 ppm (high) CO₂ concentrations because it is twice as high as current levels but still within the range of projected CO₂ concentrations by the end of this century (7). In our module, all environmental parameters other than CO₂ are held constant: the temperature in all chambers is 23°C; the relative humidity is held at 70%; and *B. rapa* plants are grown in continuous light (for more information on planting and growing conditions, see Supporting File S1: Stomatal Density Lesson). Thus, students can isolate the effects of CO₂ on the plants.

This module was developed so students can synthesize results from each lesson to gain a better understanding about how plants function and grow. In addition, when students begin their research, we ask them to consider how elevated CO₂ will ultimately impact plant fitness and the future existence of plant species. Not only is this an interesting biological question, it also has direct impact on humans, since human life depends on plants, which provide oxygen, food, clothing, shelter, and medicine.

Intended Audience

This module was developed for students in an introductory biology laboratory course at a 4-year college. However, each lesson could be adapted to general education or upper level-plant biology courses.

Required Learning Time

This module requires four-weeks, during which students meet weekly for one, three-hour laboratory. Additionally, in our curriculum students attend twelve 50-minute lectures that focus on plant biology and are complementary to laboratory material presented in these lessons. Students could also attend a weekly one-hour period to work on data analysis and collaborative writing of scientific reports related to each week's lesson.

Pre-requisite Student Knowledge

Students are required to read the background material about general plant responses to CO₂ (Supporting File S6: Pre-module Background Reading), as well as procedures associated with each week's lesson before they come to class (Supporting Files S1-S4: See the Introduction and Procedure Sections in each Lesson). This requirement familiarizes students with basic concepts, terms, and procedures. We require students to complete a pre-lesson assignment in their lab notebooks in which they record the title, lesson objective(s), working hypothesis, and summarized procedures to ensure they are

prepared for the lesson.

Pre-requisite Teacher Knowledge

To teach these lessons, teachers need a basic understanding of plant biology, compound microscopes, and spectrophotometry, as well as the ability to grow or have access to plants for student use in the classroom. Teachers can receive basic training related to these topics on various online platforms (e.g., 35, 36, 37, 38). Teachers should also be familiar with independent and paired samples t-tests if they plan for students to use statistics to analyze data.

SCIENTIFIC TEACHING THEMES

Active learning

In Class:

In contrast to passive learning, active learning involves students directly engaging in activities that shift the responsibility of learning from the instructor to the students. In this module, students work in small groups (3-4 students) throughout the entire module. During collaboration, students actively work together as they build scientific skills. In addition, students participate in hands-on laboratory activities that foster problem-solving and promote the construction of new knowledge. Also, students are responsible for actively maintaining an accurate, up-to-date lab notebook for the duration of the module.

Outside of Class:

Students perform and interpret data analyses and work with their small groups. They contribute to their lab notebook before each week's lesson by preparing a pre-lab section. Students also work on collaborative writing with peer review outside of class.

Assessment

Assessments for these modules include data gathered from five sources: (1) laboratory notebooks, (2) written lab reports, (3) student evaluation of participation of group members (Supporting File S7: Group Member Evaluation Form), (4) assignments (optional Pre-Lesson Worksheet in Supporting File S3: Foliar Protein Lesson), and (5) optional quizzes (Supporting File S8: Example Quiz).

First, laboratory notebooks are informally spot-checked to make sure that students are actively recording their activities. In addition, these notebooks are formally evaluated according to a grading rubric (Supporting File S9: Lab Notebook Grading Rubric) that assesses students on several key criteria (for criteria, see Supporting File S9: Lab Notebook Grading Rubric). Over the course of the four labs, each criterion is assessed at least two times (for details, see Supporting File S9: Lab Notebook Grading Rubric).

Second, students submit collaboratively written lab reports that are in the style of a scientific paper. Students have opportunities to submit multiple drafts, requiring both self-revision and peer-revision. Reports are graded according to a rubric (Supporting File S10: Collaborative Report Instructions and Grading Rubric). All students in a group experience being the author of a section and an editor for different sections. The grading process and author/editor model can be modified

as desired to work in any laboratory context (for details, see Supporting File S10: Collaborative Report Instructions and Grading Rubric).

Third, each student submits a self-assessment that characterizes their own participation in the group and participation levels of other group members using a standard form at the end of each module (Supporting File S7: Group Member Evaluation Form). Although research on the value of student self-assessments (SSAs) is mixed (39), there is evidence linking SSA to enhanced metacognition and learning outcomes (39) and increased student motivation (40). Fourth, assignments from specific lessons are sometimes used as formative assessments. For example, the foliar protein lab (Supporting File S3: Foliar Protein Lesson) includes an assignment that can be used as a formative assessment. Finally, quizzes can be periodically given after each lesson or at the end of the module to gauge student understanding (Supporting File S8: Example Quiz).

Inclusive teaching

This module is designed to mimic a true research experience as students apply findings from each week's lesson to inform hypotheses for the following weeks' lessons. By accurately representing the nature of science and scientific knowledge, these lessons promote problem-solving and curiosity as opposed to rote memorization. In doing so, students of all backgrounds find intellectual substance through active learning and have a chance to grow in their understanding of the nature of science, regardless of prior background.

In addition, students work in small, collaborative groups throughout the module. In these groups, students leverage each other's strengths and knowledge as they make predictions, test those predictions, gather evidence, analyze data, and communicate results. Furthermore, the module includes a breadth of activities including manipulation of live plants, conducting primary literature research, discussing/debating results, and co-writing/peer-editing the final lab report. Activities such as these that utilize multiple representations and engage students in multimodal experiences to promote inclusive learning for all races and ethnicities (41).

LESSON PLAN

Brassica rapa Growing Conditions

We purchase *B. rapa* seeds from Carolina Biological, although any standard biological supply company typically carries the variety we use (Supporting File S5: Supply List). Standard *B. rapa* Fast Plants® seeds should be planted 10-14 days before the first lesson (Table 1). We plant seeds in accordance with the Fast Plants® Quad Growing System (42) with the exception that we use four pellets of Brassica fertilizer instead of three pellets. We have found that increasing the fertility improves plant vigor under our growing conditions.

Once planted, *B. rapa* should be grown in growth chambers with a temperature of 23°C, relative humidity of 70%, and continuous light. We position the tops of the plants at approximately 10 cm below 23 Watt, compact, fluorescent lights. To test the impact of elevated atmospheric CO₂, half of the plants should be grown in 400 ± 50 ppm of CO₂ and

the other plants at 800 ± 50 ppm of CO₂. The soil and plants stay relatively moist using the wicking mats as instructed by the Fast Plants® Growing System, but we additionally water cells every two to three days if they appear dry to maintain plant vigor. This additional watering is especially important during the germination period. Approximately three days after planting, we thin plants to one per cell (Table 1).

Before class begins, determine how many total plants are needed, but plan to grow extras to account for plant mortality. For Lesson 1 (Supporting File S1: Stomatal Density Lesson), each student analyzes two leaves: one leaf from a plant grown in high CO₂ and one leaf from a plant grown in low CO₂. If the plant has four true leaves (not cotyledons), then two plants are sufficient for a group of three to four students. After the leaves are removed from the plant, the plants are discarded because they no longer have enough leaves to survive. Similarly, two to four plants are needed per group for Lesson 2 (Supporting File S2: Photosynthesis and Respiration Lesson). For Lesson 2, the aboveground parts of the plants are excised from the roots to measure photosynthetic and respiratory rates, and thus the plants cannot be used again. For Lesson 3 (Supporting File S3: Foliar Protein Lesson), only one plant grown in high concentrations of CO₂ and one plant grown in low concentrations of CO₂ are needed per student group. Only one leaf per plant is analyzed, so the plants can be used again for Lesson 4 (Supporting File S4: Growth and Allocation Lesson) as long as each plant is returned to the correct CO₂ concentration. See Table 2 for a breakdown of how many plants are typically grown for 50 groups of four students. The total number of plants needed to conduct all the lessons may seem overwhelming, but it is easy using Fast Plants®. Each tub grows 32 plants and only takes 32 cm x 20 cm of surface space! For example, we grow over 700 plants in four growth chambers.

Overview of Plant Module

The plant module is designed to answer the following question, “How will plants respond to higher concentrations of CO₂?” (Supporting Files S1-S4). The first lesson (Supporting File S1: Stomatal Density Lesson) allows students to visualize and count stomata on the underside of leaves using compound microscopes. Each student analyzes two leaves, one from a plant grown in 400 ppm of atmospheric CO₂ and one from a plant grown in 800 ppm of atmospheric CO₂. Then, students calculate stomatal density for each plant and enter their data into the course database. Students statistically analyze course results using an independent samples t-test. Based on their findings, students predict how CO₂ concentrations will impact photosynthetic rates in Lesson 2.

In Lesson 2 (Supporting File S2: Photosynthesis and Respiration Lesson) students measure photosynthetic and respiratory rates of *B. rapa* in ambient conditions of CO₂, after being grown under high or low CO₂ concentrations. Plants are cut at the soil level so that their mass can be measured. Plants are then placed into a chamber connected to a CO₂ gas sensor that measures CO₂ concentrations in the chamber. The chambers with the cut plants are placed under a fluorescent lamp, and CO₂ concentrations in the chamber are measured for 15 minutes. Next, the chamber is wrapped in aluminum foil, and plants acclimate to the darkness for a 5-minute period.

CO₂ concentrations are again measured in the chamber for 15 minutes. Students use the slope of the regression line between CO₂ and time to estimate net photosynthetic and respiratory rates (ppm/min). Photosynthetic rates and respiratory rates are then standardized on a per-mg basis [(ppm/min)/mg]. As in Lesson 1, students enter data into a course database and statistically analyze the results.

The third lesson (Supporting File S3: Foliar Protein Lesson) allows students to measure water-soluble protein concentrations from the leaves of *B. rapa* grown in the high and low atmospheric CO₂ concentrations. In this lesson, we assume the soluble protein present is predominantly comprised of photosynthetic enzymes like Rubisco, which binds CO₂ from the atmosphere. Protein is extracted from leaves and combined with Bio-Rad protein assay dye. The concentration of the protein is then estimated using a spectrophotometer and a bovine serum albumin standard curve. As in the previous lessons, students enter data into a course database and statistically analyze the results.

The fourth and final lesson (Supporting File S4: Growth and Allocation Lesson) enables students to analyze growth and patterns of resource allocation of *B. rapa* grown in the different CO₂ concentrations. Each student analyzes one plant from each of the CO₂ treatments. Plants are removed from their cells, and roots are submerged in a tube of water and swirled around until the soil is removed. Students then measure total plant mass and partial plant mass of various other parts (e.g., root, shoot, and fruit). In addition, large seedpods are removed and dissected to count the number of mature, viable seeds. Once data from these variables are gathered, students calculate the root:shoot ratio and reproductive:vegetative ratio. These ratios indicate if the plant allocated photosynthate (i.e., carbon-based compounds) differently to plant structures. As before, students enter data into a course database and statistically analyze the results.

Surprisingly, we have found that *B. rapa* are unresponsive to elevated CO₂ for all responses measured (Table 3), which does not support the majority of published research findings (e.g., 8, 12, 14, 15, 16, 23). We use this opportunity to discuss the importance of anomalous and negative results. Most students assume that future elevated CO₂ conditions will be beneficial for plants. However, after this module students realize that responses can be species specific. We then brainstorm hypotheses about why *B. rapa* may not follow the general trend of other C3 plants. One common hypothesis is that *B. rapa* (Fast Plants®) have been bred to grow at very high rates and that they may have reached a physiological maximum and cannot photosynthesize or grow at faster rates. Recently, we have also begun to explore different species (i.e., peppers) in our courses that do respond to elevated CO₂ (43). Then we conduct small and large group discussions comparing results across species and brainstorm why some species respond to elevated CO₂ while others do not.

Miscellaneous: Data File, Statistics, Lab Notebooks, and Writing

This module was designed for introductory-level biology students who want to be biology majors and/or want to

pursue professional, health-related fields. In our inquiry-based labs for introductory biology, we emphasize the importance of supporting results with statistical evidence (i.e., claim - evidence - reasoning). Therefore, students use t-tests every week during this module to analyze data and interpret the results. To gather course data, we create a course database in which students across all sections enter their data. Then, the data are checked for errors and extreme outliers (two orders of magnitude or more) before the final data set is posted for data analysis.

In our introductory biology lab course, students keep a lab notebook with the following required information for each lesson: date, experiment title, group members' names, purpose of experiment, working hypothesis, data, calculations, statistical hypotheses (H₀ and H_a), t-test output, figures and tables, results, and conclusions. At the end of the module, we assess the lab notebook using a grading rubric that consists of selected items from the required list assigned to each week's lab (i.e., "spot check"; Supporting File S9: Lab Notebook Grading Rubric). This approach allows us to give feedback to students as they learn these skills without grading the entire lab notebook.

In addition, students gain experience in scientific writing and peer-reviewing in this module by authoring a report in the style of a scientific journal. The writing assignments are designed to simulate how a group of collaborators might write and edit each other's work. During the first three lessons, a different section of a scientific paper (e.g., methods, results, etc.) is emphasized and discussed so that students can learn about that particular section. Students take turns writing and peer-reviewing these sections of the report until week four. At this point, students collaborate to make a final (and complete) report. In the final report, students divide the paper into sections, and each student serves as the lead author on at least one section. All students in the group also serve as editors of the other sections of the report. We feel this strategy helps ensure that everyone is involved in the writing process. Peer editors provide feedback using the comments feature in word processing software (e.g., Microsoft Word, Google Docs). Peer edits are retained, printed, and graded along with the final version of each section and completed report. All graded items are graded according to grading rubrics (Supporting File S10: Collaborative Report Instructions and Rubric).

TEACHING DISCUSSION

These four lessons were designed to help students understand how anatomical (stomata), physiological (photosynthesis/respiration), and biochemical attributes (protein concentration) of plants interact and allow plants to survive, grow, and reproduce. We felt that investigating the impact of atmospheric concentrations of CO₂ on plants provided an environmental context that many students would find relevant and relatable to societal concerns. In addition to content knowledge, this module was designed to develop students' basic scientific skills through authentic laboratory experiences. Specific skills that were emphasized included keeping lab notebooks, performing statistical analysis, writing scientific papers, and group collaboration.

Student Reactions to Lessons and Assessments

Our previous experiences indicate that students do not typically enjoy keeping a lab notebook as it takes considerable time. However, we felt this was an important scientific skill as professional labs require scientists to keep detailed records and most health-related professions require accurate and detailed record-keeping. Therefore, to minimize student time on lab notebooks out of class, we emphasized that students get as much done as possible during lab and required that students had recorded all their group's data in their lab notebooks before they could leave lab. We have found that many students were not effectively using class time and did not record data as they worked because they wanted their lab notebooks to be "perfectly organized," which is unrealistic and not an authentic scientific practice.

Additionally, students had mixed reactions to statistical analysis of data. At the beginning of the course, many students said that statistical analysis was confusing. However, we typically observed that most students easily conducted and interpreted t-tests after a few weeks of practice. Some students appreciated the use of probability and analytical evidence in interpreting and providing evidence for their results. Moreover, students who have gone on to graduate school or research have told us that they were better prepared than their peers to conduct research and succeed in graduate-level courses in part because of their statistical skills.

As part of our assessment, we solicited feedback from teaching assistants (TAs) as well as students. Our TAs were undergraduates who had taken the course previously. Feedback from TAs indicated the plant module effectively helped students achieve the desired learning goals. TAs noted that the related lessons gave students a broader understanding of plants, and according to one TA, brought "a simple topic... to life before the students' eyes." TAs also indicated the modular approach facilitated students' making conceptual connections. For example, students learned to evaluate *B. rapa*'s resource allocation based on varying CO₂ levels and previously measured attributes (e.g., stomatal density) and processes (e.g., net photosynthesis). One TA stated, "I think students engaged in the growth and resource allocation lab... some were asking me good questions about *Brassica rapa*."

TAs generally found that students had positive attitudes about this module. For example, TAs reported that students were "fascinated with how something so small [stomata] could be found and evaluated during lab." Students also appreciated the practical nature of the lab as they investigated how "simple plants are alive and constantly doing important chemical reactions." Furthermore, TAs stated that students were engaged because of the environmental relevance of the topic. According to one TA, students "evaluate the effect of increased levels of carbon dioxide, which is becoming a world problem. [Students] get to help research the effects." Students seemed to appreciate the opportunity to do research on an important environmental topic.

TAs advised that the final report is an important component. Students learned to ask questions, formulate hypotheses, gather/analyze data, predict outcomes, and display results. However, students also had to think deeply about these components by writing and reviewing the final report. One TA explained how writing "requires students to understand."

Moreover, at the end of the module, students will be able to connect findings and ideas from all lessons to get a bigger picture of how plants work.

Adaptability of Lessons

While this module works well in our current setup, we recognize the ability to grow plants in elevated CO₂ conditions will likely be limited at many institutions. Therefore, analyzing different independent variables (e.g., soil fertility, light intensity, soil contaminants, etc.) in your classrooms, guided by both interest and available equipment, is encouraged. Similarly, we use *Brassica rapa* Fast Plants® for our research because they mature from seed to flowering in about two weeks and take very little space. However, any small, relatively fast-growing plant would work, if the space and resources needed to grow the plant species are available. For instance, following student interest, we recently substituted *B. rapa* with a hot pepper plant (*Capsicum annuum* 'Poinsettia') in this module and had an interesting module of experiments where we could compare responses of a different species to responses of *B. rapa*.

In our course, students collaborate in a group to write a group lab report. However, depending on course goals or instructor preferences, each student could instead write an individual lab report. One model we have used in the past is for one student per group to choose one of the four lessons to write his/her report. This model also works well because students have input about what they will write. Additionally, the papers are then turned in over a four-week period, helping the instructor keep up with grading.

Finally, each lesson could easily be taught in isolation. The ability for students to complete and understand each lesson is not contingent on completing any other lesson beforehand.

SUPPORTING MATERIALS

- S1. Plants and CO₂-Lesson 1: Stomatal Morphology and Density: Student Lab Manual Pages (.docx)
- S2. Plants and CO₂-Lesson 2: Photosynthesis and Respiration: Student Lab Manual Pages (.docx)
- S3. Plants and CO₂-Lesson 3: Foliar Protein Concentrations: Student Lab Manual Pages (.docx)
- S4. Plants and CO₂-Lesson 4: Growth and Resource Allocation: Student Lab Manual Pages (.docx)
- S5. Plants and CO₂-Supply List per Lesson (.docx)
- S6: Plants and CO₂-Pre-module Background Reading: Plant Responses to CO₂ (.docx)
- S7. Plants and CO₂-Group Member Evaluation Form (.docx)
- S8. Plants and CO₂-Example Quiz (.docx)
- S9. Plants and CO₂-Lab Notebook Sample Grading Rubric (.docx)
- S10. Plants and CO₂-Collaborative Report Instructions and Grading Rubric (.docx)

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Table 1. Plants and CO₂ - Teaching Timeline - This table is intended to outline the timeline of the four-week set of four lessons. The instructor can keep track of *Brassica rapa* care protocol, as well as notes for each weekly lesson.

Week	Notes	Estimated Time	Supporting Files
Before Lesson 1			
<i>Brassica</i> care for Lessons 1, 3, and 4	10 -14 days before the stomatal density lesson, plant <i>Brassica</i> seeds and set-up growth chamber	Time depends on plant number needed. I can prepare 32 plants in 20 minutes.	<ul style="list-style-type: none"> Fast Plants® Quad Growing Protocol (http://www.fastplants.org/pdf/grow/quad_protocol.pdf) Supporting File S5: Supply List
<i>Brassica</i> care for Lessons 1, 3 and 4	Approximately 3 days after planting, thin seedlings to one plant per cell so plant density does not impact <i>Brassica</i> growth	Approximately 1 hour for 200 cells	
<i>Brassica</i> care for Lesson 2	11-13 days before the photosynthesis/respiration lesson, plant additional <i>Brassica</i> seeds. This separate planting date helps ensure that plants are developmentally young and thus have negative net photosynthetic rates (meaning photosynthetic rates are greater than respiratory rates). More mature plants with flowers or seeds frequently have positive net photosynthetic rates, which is conceptually more difficult for students. Note: We typically do not thin plants after seed germination for this lesson to achieve higher photosynthetic rates		<ul style="list-style-type: none"> Fast Plants® Quad Growing Protocol (http://www.fastplants.org/pdf/grow/quad_protocol.pdf) Supporting File S5: Supply List
Week of Lesson 1			
Lesson 1: Stomatal Density	Students remove leaves from plants for this experiment.	One 3-hour lab period	<ul style="list-style-type: none"> Supporting File S1: Stomatal Density Lesson Supporting File S5: Supply List
Week of Lesson 2			
<i>Brassica</i> care for Lessons 3 and 4	15-17 days after planting seeds, use a paintbrush to transfer pollen among plants. Pollination is necessary for fruit/seed production, which will be measured in Lesson 4.	Approximately 20 - 30 minutes per 100 plants	Wisconsin Fast Plants® Growing Instructions (42)
Lesson 2: Photosynthesis and Respiration	Plants need to be 11-13 days old for CO ₂ intake rates (photosynthesis) to be greater than CO ₂ expulsion rates (respiration). The shoot (above-ground plant parts) must be cut off at the soil level and immediately placed in the Vernier respiration chamber to analyze [CO ₂] (Supporting File S5: Supply List)	One 3-hour lab period	<ul style="list-style-type: none"> Supporting File S2: Photosynthesis and Respiration Lesson Supporting File S5: Supply List
Week of Lesson 3			
Lesson 3: Foliar Protein Concentrations	Only part of one leaf is needed for analysis.	One 3-hour lab period	<ul style="list-style-type: none"> Supporting File S3: Foliar Protein Lesson Supporting File S5: Supply List
Week of Lesson 4			
Lesson 4: Growth and Resource Allocation	Students dissect and measure mature plants.	One 3-hour lab period	<ul style="list-style-type: none"> Supporting File S4: Growth and Allocation Lesson Supporting File S5: Supply List

Table 2. Plants and CO₂- Suggested number of plants for 200 students, divided into 50 groups of four students each. This table is a guide to help the instructor determine how many plants will be needed for each lesson, depending on how many students and/or groups are participating. For each lesson, half of the number of plants indicated should be grown in the low CO₂ conditions and half of the plants should be grown in high CO₂ conditions.

Lesson	Number of Plants	When to Plant Seeds	Running Total Number of Plants for Module
1: Stomatal Density (Supporting File S1)	50 groups X 2 plants* = 100 + 64 extras = 164 plants	10-14 days before Lesson 1	164
2: Photosynthesis and Respiration (Supporting File S2)	50 groups X 2 plants* = 100 + 64 extras = 164 plants	11-13 days before Lesson 2	328
3: Foliar Protein Concentrations (Supporting File S3)	50 groups X 2 plants* = 100 + 64 extras = 164 plants	10-14 days before Lesson 1	492
4: Growth and Resource Allocation (Supporting File S4)	200 students X 2 plants* = 400 plants – 164 plants that are reused from Lesson 3 = 236 plants	10-14 days before Lesson 1	728
<i>*Each week, students or student groups analyze two plants: one plant grown in low CO₂ conditions and one plant grown in high CO₂ conditions.</i>			

Table 3. Plants and CO₂- Response of *Brassica rapa* (Fast Plants®) to low (400 ppm) and high (800 ppm) concentrations of atmospheric CO₂. This table shows an example of our results from one cohort of students in our introductory course. Results can vary by plant species and growing conditions used.

Lesson	Dependent Variable	# of Plants Analyzed	400 ppm CO ₂ Mean ± SEM	800 ppm CO ₂ Mean ± SEM	*p-value
Lesson 1 (Supporting File S1)	Stomatal Density (stomata/mm ²)	110	175.4 ± 12.1	187.1 ± 12.7	0.096
Lesson 2 (Supporting File S2)	Net Photosynthetic Rate [ppm/min)/mg]	114	0.0051 ± 0.002	0.0056 ± 0.002	0.862
	Respiratory Rate [ppm/min)/mg]	114	0.0315 ± 0.002	0.0335 ± 0.002	0.505
Lesson 3 (Supporting File S3)	Foliar Protein Concentration (mg protein/mg leaf)	68	0.0064 ± 0.001	0.0059 ± 0.001	0.501
Lesson 4 (Supporting File S4)	Total Plant Mass (mg)	306	1602 ± 56.7	1550 ± 54.6	0.508
	Fruit Mass (mg)	306	375 ± 27.1	427 ± 28.3	0.179
	Root:Shoot	306	0.44 ± 0.18	0.26 ± 0.02	0.324
<i>*p-values were generated by independent samples t-tests for all lessons except Lesson 3, Foliar Protein Concentration, which was generated by a paired-sample t-test.</i>					